

Figure 1. A plot showing the dependence of k_{obsd} on the acetic acid concentration for the hydrolysis of benzaldehyde *O*-ethyl *O*-trifluoroethyl acetal at 25 °C and $\mu = 0.5$ (KCl). $k_{\text{obsd}}/[\text{H}^+]$ is plotted vs. $[\text{HOAc}]/[\text{H}^+]$ (cf. eq 2 discussion); \circ points are at pH 4.65 (± 0.05), $[\text{HOAc}]/[\text{OAc}] = 1.0$ and \square points are at pH 4.98 (± 0.06), $[\text{HOAc}]/[\text{OAc}] = 0.50$. The slope = $k_{\text{ac}} = 7.84 \times 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$, the intercept = $k_{\text{H}^+} = 12.3 \text{ M}^{-1} \text{ s}^{-1}$, and the correlation coefficient is 0.996.

Mixed acetals, e.g., benzaldehyde *O*-ethyl *O*-2-chloroethyl acetal, were prepared by an adaptation of analogous literature procedures.^{3b,10} Typically, 0.6 mol of the diethyl acetal of benzaldehyde was mixed with 0.6 mol of thionyl chloride. After standing for 30 min, the volatile side products and excess thionyl chloride were removed by vacuum distillation at 30 Torr to a pot temperature of 100 °C. Vacuum distillation at ca. 1 Torr produced an essentially quantitative yield of the α -chloro ether, α -chloro- α -ethoxytoluene (bp 72–74 °C, 1 mm). Because of the high reactivity and toxicity of α -chloro ethers, the distillate was collected in 0.1-mol batches in round-bottomed flasks containing 0.1 mol of the appropriate alcohol, 0.1 mol of NaH, and 100 mL of DMF. A special fraction cutter was used which allowed the addition of the α -chloro ether to the stirred DMF solution to be made at room pressure without interrupting the vacuum distillation. Following a basic aqueous workup procedure, 50–75% yields of vacuum-distilled *O*-ethyl *O*-alkyl acetals were obtained and ascertained to be pure by ¹H NMR analysis.

Kinetic Method. Reactions were monitored at the absorption maximum of the benzaldehyde produced by hydrolysis, using either a Beckman Kintrac VII or a Beckman Recording DU with a specially constructed constant-temperature bath positioned between the monochromator and the phototube housing. (In the latter case the temperature was maintained at 25.00 ± 0.02 °C.)¹¹ Reaction conditions were pseudo first order and plots of $\log(A_\infty - A_t)$ against time were linear for at least 3 half-lives of reaction time. Rate constants were calculated from these graphs and also by a computer program which fitted data for the first 3 half-lives to the best first-order curve of absorbance vs. time, using A_∞ as a variable.¹² Typically, the A_∞ values calculated by the computer program agreed with the experimentally observed values, except for some of the very slow reactions where it was inconvenient to wait for the absorbance to reach A_∞ (particularly in phosphate buffers, where the experimental A_∞ decreased with time, associated with the observed growth of organisms in the cuvette solution).

Second-order rate constants were calculated from the observed rate constants according to

$$k_{\text{obsd}} = k_{\text{H}^+}[\text{H}^+] + k_{\text{HA}}[\text{HA}] \quad (2)$$

where $[\text{H}^+]$ is antilog ($-\text{pH}$), as measured by a Beckman Model 4500 pH meter using a combination glass electrode. In most cases it was possible to measure k_{H^+} in 10^{-3} – 10^{-1} N HCl solutions where the last term of eq 2 is negligible. For buffer solutions, plots of $k_{\text{obsd}}/[\text{H}^+]$ against $[\text{HA}]/[\text{H}^+]$ produced straight lines (cf. Figure 1) whose slope = k_{HA} and intercept = k_{H^+} . (In no cases were pH-independent terms detectable in eq 2.) Values of k_{H^+} obtained from the intercept were identical with those obtained in dilute HCl solutions. Slopes of plots

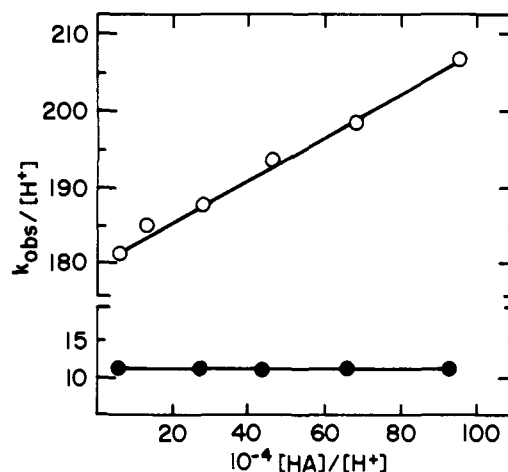


Figure 2. A same-scale superimposition of Figure 1-type plots for the hydrolyses of diethyl acetals of benzaldehyde (\circ , slope = $k_{\text{cac}} = 3.01 \times 10^{-5} \text{ M}^{-1} \text{ s}^{-1}$, intercept = $k_{\text{H}^+} = 180 \text{ M}^{-1} \text{ s}^{-1}$, corr = 0.996) and *m*-chlorobenzaldehyde (\bullet , slope = $k_{\text{cac}} = 0$, intercept = $k_{\text{H}^+} = 12 \text{ M}^{-1} \text{ s}^{-1}$) in cacodylic acid buffers, $\mu = 0.5$ (KCl), 25 °C, $[\text{HA}]/[\text{A}^-] = 2.0$, [cacodylic acid] varied from 0.08 to 0.80 M.

such as Figure 1 were independent of buffer ratio. The advantage of plotting $k_{\text{obsd}}/[\text{H}^+]$ against $[\text{HA}]/[\text{H}^+]$ is that it takes into account the slight dependence of pH on the buffer concentration which arises because of the small specific salt effect on $\text{p}K_{\text{HA}}$. Values of $\text{p}K_{\text{HA}}$ which are cited are values at zero buffer concentration and 0.5 N KCl (obtained by extrapolation of a plot of $\text{p}K_{\text{HA}}$ against $[\text{buffer}]$ to $[\text{buffer}] = 0$).

Results

Hemiacetal Buildup. Most of the kinetic studies in buffers were at a sufficiently high pH to ensure that the hemiacetal hydrolyzed quickly to benzaldehyde via a base-catalyzed pathway, thus assuring that the rate-determining step was truly acetal hydrolysis and not hemiacetal hydrolysis.^{4a} In dilute HCl solution, the first 10–20% reaction was not first order because of the time required to produce a pseudo-steady-state amount of hemiacetal; however, the absorbance–time data exhibited first-order behavior throughout the rest of the measurable reaction and it has been shown that the rate measured for this portion of the reaction is essentially that of acetal hydrolysis.^{4a}

General Acid Catalysis. Plots such as Figure 1 show that acetals of benzaldehydes hydrolyze by a process that exhibits a rate dependence on buffer concentration. These rates can be analyzed according to eq 2 as described; however, two concerns remain.

(1) It must be demonstrated that it is the acid component of the buffer which functions as catalyst. This can be unambiguously demonstrated by showing that the slopes of plots such as Figure 1 are independent of buffer ratio; such was the case in all our experiments (cf. Figure 2).

(2) For cases where the slopes of plots such as Figure 1 are shallow, there must be some assurance that “catalysis” is not merely a specific salt effect.¹³ One of the least ambiguous “checks” is that used in this study. A reaction may be shown to exhibit a rate dependence on buffer concentration well above that for some closely related reaction. That is, the closely related reaction can be used to “calibrate” the specific salt effect. For those cases where no rate dependence on $[\text{buffer}]$ existed (e.g., substrates with large α studied in the more acidic buffers), plots such as Figure 1 had a slope of zero. Thus, there appears to be no detectable specific salt effect in aqueous buffer solutions. This point is not trivial and was not easily resolved. For example, a portion of this study was intended to investigate the effects of mixed solvents on general acid catalysis; however, even in 20% dioxane–water or 20% Me_2SO –water mixtures,

Table I. Second-Order Rate Constants for the General-Acid-Catalyzed Hydrolyses of Dialkyl Acetals of Para-Substituted Benzaldehydes in Water at 25 °C, $\mu = 0.5$ (KCl)^a

OR	10 ⁵ k _{mp} ^b	10 ⁴ k _p ^c	10 ⁴ k _{mes} ^d	10 ⁴ k _{cac} ^e	10 ³ k _{ac} ^f	10 ² k _F ^g	k _{H+}	α ^h	corr ⁱ
OCH ₂ CH ₃	308	88.5		X = Me ₂ N 251			30,400	0.81	0.999 96
OCH ₂ CH ₃	11.4		5.83	X = CH ₃ O 7.64			2 450	0.85	0.999 90
OCH ₂ CH ₂ OCH ₃			0.681	1.56	3.10		262	0.83	0.999 90
OCH ₂ CH ₂ Cl	0.743	0.367		0.942	1.02	1.12	61.8	0.78	0.9990
OCH ₂ CH ₃			1.33	X = CH ₃ 2.19			725	0.86	0.999 991
OCH ₂ CH ₃			0.307	X = H 2.95	0.858		169	0.87	0.9994
OCH ₂ CH ₂ OCH ₃				0.0359	0.079		16.8	0.88	0.999 95
OCH ₂ CH ₂ Cl					0.0362		3.98	0.84	
pK' _{HA} ^j	7.41	6.59	6.18	6.19	4.59	3.53			

^a Mean values of replicate determinations at different buffer ratios; the standard deviation typically is ± 2 –5% for k_{H+} and ± 5 –10% for k_{HA} values. ^b mp = CH₃PO₃H⁻. ^c p = H₂PO₄⁻. ^d mes = 2-(*N*-morpholino)ethanesulfonic acid, a Calbiochem buffer. ^e cac = (CH₃)₂AsO₂H. ^f ac = CH₃CO₂H. ^g F = HCO₂H. ^h α = Brønsted α. ⁱ corr = correlation coefficient. ^j pK'_{HA} = pK_a of the buffer acid HA at $\mu = 0.5$ and zero buffer concentration.

Table II. Second-Order Rate Constants for the General-Acid-Catalyzed Hydrolyses of Benzaldehyde *O*-Ethyl *O*-Alkyl Acetals in Water at 25 °C, $\mu = 0.5$ (KCl)^a

OR	10 ⁵ k _{mes} ^b	10 ⁵ k _{cac} ^c	10 ⁴ k _{ac} ^d	10 ³ k _F ^e	k _{H+} ^h	α ^f	corr ^g
OCH ₂ CH ₃	3.07	2.95	8.58		169	0.87	0.9994
OCH ₃	1.18	1.80	5.80		122	0.89	0.999 94
OCH ₂ CH ₂ OCH ₃		2.81	5.26	8.52	82.6	0.86	0.991
OCH ₂ CH ₂ Cl		2.48	5.02	3.54	50.5	0.83	0.999 95
OCH ₂ C≡CH		3.97	5.86	6.60	37.3	0.79	0.9994
OCH ₂ CF ₃		6.78	7.82	4.47	12.6	0.70	0.999 95

^a Mean values of replicate determinations at different buffer ratios; the standard deviation typically is ± 2 –5% for k_{H+} and ± 5 –10% for k_{HA} values. ^b mes = 2-(*N*-morpholino)ethanesulfonic acid (a Calbiochem buffer). ^c cac = (CH₃)₂AsO₂H. ^d ac = CH₃CO₂H. ^e F = HCO₂H. ^f α = Brønsted α. ^g corr = correlation coefficient. ^h ρ* = -1.06 (corr = 0.992).

a significant negative salt effect was observed. (Plots such as Figure 2 for mixed solvent systems produced a line of significant negative slope for the hydrolysis of *m*-chlorobenzaldehyde diethyl acetal). Such behavior was eliminated by the use of KCl as the common salt and by the use of a totally aqueous solvent. The decision to work at $\mu = 0.5$ was dictated by the marginal solubility of some of the acetals at higher ionic strengths.

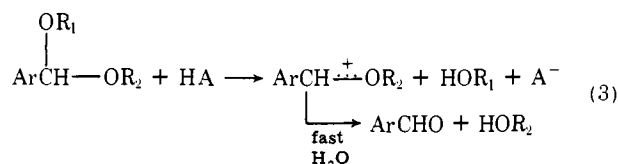
The substance showing the largest α (the least general acid catalysis) is the diethyl acetal of benzaldehyde. Figure 2 displays a comparison of the rate dependence on buffer concentration for the diethyl acetals of benzaldehyde and *m*-chlorobenzaldehyde. The slopes of the lines for the two substances are significantly different and the difference in slope is dependent upon the buffer acid; however, in both cacodylic acid and acetic acid buffers the slope of the line for *m*-chlorobenzaldehyde diethyl acetal is essentially zero. Therefore, we are confident that the k_{HA} calculated for benzaldehyde diethyl acetal from the slopes of plots such as Figure 2 is truly the rate constant for that portion of the hydrolysis catalyzed by the buffer acid as defined by eq 2.

The Brønsted plots constructed from the data of Tables I and II show no detectable dependence of k_{HA} on charge type; i.e., the points for buffers of the type RPO₃H⁻ and the zwitterionic MES buffer lie on the line established by the proton and RCO₂H points. Thus the type of charge effect reported

by Kresge²¹ for the general-acid-catalyzed hydrolyses of vinyl ethers is not significant for the hydrolyses of acetals of benzaldehydes.

Discussion

General Acid Catalysis. The observation of general acid catalysis for the variety of acetals of benzaldehydes listed in Tables I and II requires that eq 1 be incorrect for these hydrolyses. The mechanism formerly proposed^{2,3} for the "special cases" is eq 3.



It is instructive to consider this mechanism for the present series because it affords the first opportunity to observe the transition from a "special case" acetal (where general acid catalysis is observed) to a "typical" acetal (where general acid catalysis is not observed). It is particularly informative to analyze this system using the Jencks-O'Ferrall approach^{5,14} as described by Figure 3. Taking the diethyl acetal of benzaldehyde as the reference compound, the large Brønsted α (0.87,

Table III. Second-Order Rate Constants for the Hydrolyses of Diethyl Acetals of Substituted Benzaldehydes in Water and Dioxane-Water Mixtures at 25 °C, $\mu = 0.5$ (KCl)

substituent	$k_{H^+}^a$		
	water	20% dioxane- water	50% dioxane- water
<i>p</i> -NO ₂	0.519	0.221	0.287
<i>m</i> -NO ₂	0.952	0.393	0.0402
<i>m</i> -Cl	11.5	5.03	0.466
<i>m</i> -OCH ₃	82.8	43.7	8.15
H	169	94.2	15.8
<i>m</i> -CH ₃	233	142	22.7
<i>p</i> -CH ₃	725	415	81.7
<i>p</i> -OCH ₃	2 450	1 580	491
<i>p</i> -NMe ₂	30 400	44 700	21 100
ρ^b	-3.16 (0.999)	-3.35 (0.999)	-3.58 (0.996)
ρ_r^b	-0.29	-0.60	-0.93

^a $k_{H^+} = k_{\text{obsd}}/[H^+]$. Values listed are means of replicate measurements. k_{H^+} is independent of pH'. Average deviations from the means of replicate determinations of k_{H^+} were $< \pm 5\%$. ^b Calculated after the method of ref 6. Values in parentheses are correlation coefficients for the line generated by meta and electron-withdrawing substituents. σ values taken from J. Hine, "Structural Effects on Equilibria in Organic Chemistry", Wiley, New York, 1975.

Table I) and Hammett ρ (-3.2, Table III) locate the transition state in the lower left quadrant as shown. The following observations are derived from the α and ρ values in Tables I-III.

(a) Electron-donating groups in the Ar moiety lower the entire bottom side of the diagram in Figure 3, which causes a Hammond motion along the reaction coordinate and movement orthogonal to the reaction coordinate which sum to move the transition state in the direction designated by ② in Figure 3. This conceptual argument is corroborated by the α values of 0.87, 0.86, and 0.81 for the diethyl acetals of benzaldehyde, *p*-methyl-, *p*-methoxy-, and *p*-dimethylaminobenzaldehyde. Note that the effect of electron-withdrawing groups in the Ar moiety is just the opposite, as indicated by the motion designated ①. This is consistent with our observation that the diethyl acetal of *m*-chlorobenzaldehyde did not exhibit catalysis in the same buffers as did the diethyl acetal of benzaldehyde (i.e., α for the former must be greater than α for the latter). From the data in Table I, $\partial\alpha/\partial\sigma = 0.073$ (correlation coefficient = 0.999), which simply quantitates the observation that α is only slightly sensitive to the extent of electron donation from the aryl moiety. From this Cordes-type relationship,¹⁵ α for the diethyl acetal of *m*-chlorobenzaldehyde may be calculated to be 0.90; this is consistent with our lack of observing even a 10% rate enhancement in 1 M cacodylic acid buffers, which requires $\alpha \geq 0.91$.¹⁶

(b) As the catalyzing acid HA becomes weaker, the entire left side rises, causing Hammond motion along the reaction coordinate and movement orthogonal to the reaction coordinate which sum to move the transition state in the general direction designated by ③ in Figure 3. This is corroborated by the observation that α decreases with σ , which is equivalent to saying that ρ decreases as pK_{HA} increases. Lack of much horizontal motion in ③ is consistent with the observation that the k_{H^+} point lies on the line of the Brønsted plot. As will be elaborated further in another context, it is interesting to note that $(\partial\rho/\partial pK_{HA}) \neq 0$ requires that at some point a reversal of substituent effect occur; i.e., ρ must become positive when the catalyzing acid is sufficiently weak. In the present case, pK_{HA} would have to be impossibly large to observe this reversal of sign (larger than pK_{H_2O}), but this is simply a result of a large negative ρ for k_{H^+} and a rather small $\partial\rho/\partial pK_{HA}$.

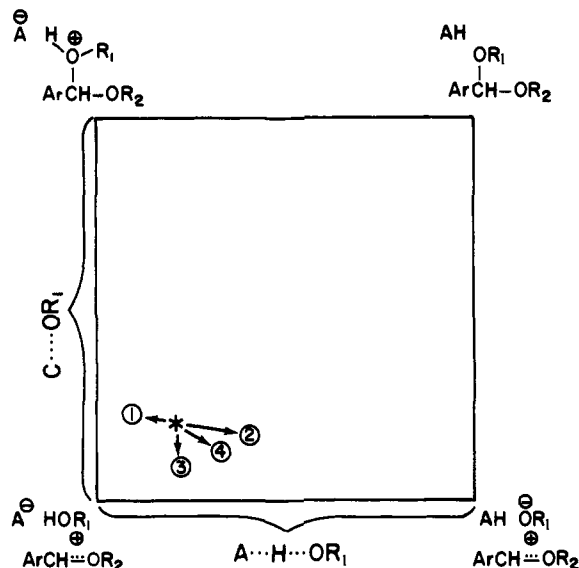
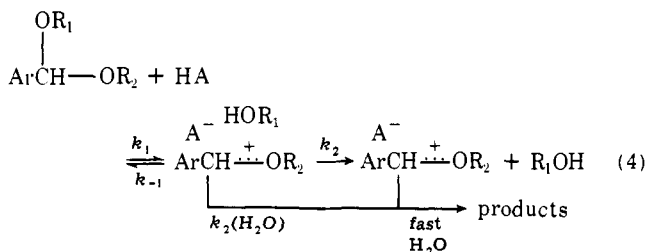


Figure 3. A Jencks-O'Farrell diagram for the hydrolysis of acetals of benzaldehydes according to eq 3. The location of the transition state for the proton-catalyzed hydrolysis of benzaldehyde diethyl acetal is located by an asterisk. The transition-state motion imparted by electron-withdrawing groups in the Ar and R₂ moieties is shown by ①, that of electron-donating groups in Ar and R₂ by ②, that of weaker catalyzing acids HA by ③, and that of electron-withdrawing groups in R₁ by ④.

(c) As R₁ becomes electron withdrawing (relative to ethyl), the upper left corner rises and the lower right corner lowers, resulting in a marked movement of the transition state in a direction orthogonal to the reaction coordinate, as designated by ④. This is corroborated by the marked decrease in α shown in Table II, $\alpha = 0.87$ for R₁ = CH₂CH₃ through $\alpha = 0.70$ for R₁ = CH₂CF₃. We have no evidence supporting the vertical motion shown by ④; the extent of this motion depends upon the direction of the reaction coordinate—we believe most of the motion shown by ④ to be horizontal rather than vertical because the reaction coordinate is probably biased toward the upper left corner for R₁ = CH₂CH₃. The $\partial\alpha/\partial\sigma^*$ correlation for this series is nonlinear, as shown in Figure 4. There are two possible explanations for the decreasing slope in moving from R₁ = CH₂CF₃ to R₁ = CH₂CH₃ on the plot of α vs. σ^* . First, it may be that a large energy barrier exists as the lower side of Figure 3 is approached, resulting in a "leveling off" of the $\partial\alpha/\partial\sigma^*$ curve. The nature of such a barrier should be a function of carboxonium ion stability; i.e., it should be affected by the shape/depth of the potential energy well in the lower left corner. The second explanation is that the "leveling off" is caused by a change in rate-determining step. An attractive mechanism is given by eq 4. As R₁ becomes more electron



withdrawing, R₁OH becomes much less nucleophilic, which decreases v_{-1} relative to v_2 ; when $v_2 > v_{-1}$, v_1 is rate determining (as for R₁ = CH₂CF₃); when $v_{-1} > v_2$, v_2 is rate determining (as for R₁ = CH₂CH₃). The presence of the conjugate base of the catalyzing acid in the transition state for the rate-limiting diffusional separation process is responsible for the slight catalysis observed ($\alpha \approx 0.88$). In the reverse direction, this amounts to a "preassociation" general-base-catalyzed mechanism.¹⁷

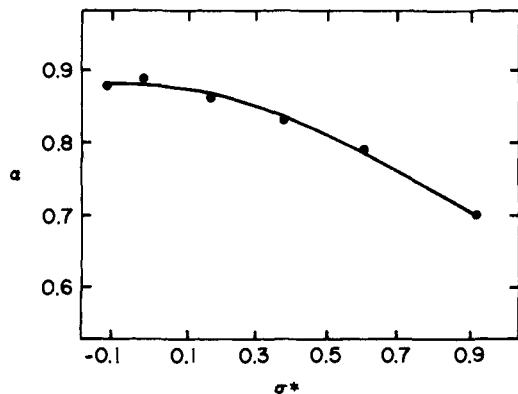


Figure 4. A plot of the Brønsted α vs. σ^* for the general-acid-catalyzed hydrolysis of benzaldehyde *O*-ethyl *O*-alkyl acetals.

At the present time we have no evidence regarding the role of water in eq 4; it is possible that, as the carboxonium ion becomes more reactive and/or R_1OH becomes more nucleophilic, the rate-determining step involves nucleophilic attack by the water. (We are beginning solvent and secondary isotope effect studies to answer this point.) For the sake of the present discussion, no distinction is made between k_2 and $k_2(H_2O)$; both are lumped together as k_2 processes.

Although the alternative explanations just presented are difficult to distinguish experimentally, several observations suggest that k_2 processes may be rate determining in some instances. Values of k_{H^+} for the series in Table II vary by only 15-fold while α changes from 0.70 to 0.87, whereas for the diethyl acetals of substituted benzaldehydes (Table I) values of k_{H^+} vary by 3000 while α changes only from 0.81 to 0.9. With reference to Figure 3, it is not clear why the ① and ② changes have such a much smaller effect on α than ④. Measured by any standard (reactivity, σ , σ^* , etc.) this is the case: the major effect of ① and ② change is perpendicular to the diagram (the energy coordinate) whereas the major effect of ④ change is along the $A \cdots H \cdots OR_1$ axis (horizontal motion on the diagram). This is most easily understood if the diethyl acetal of benzaldehyde hydrolyzes via rate-controlling diffusional separation, as per eq 4, and the rather subtle change in α which accompanies the 3000-fold change in reactivity on going from the diethyl acetal of benzaldehyde to the diethyl acetal of *p*-dimethylaminobenzaldehyde arises from a gradual change in rate-determining step (as v_{-1} decreases relative to v_2 in eq 4). That is, for rate-limiting diffusional separation, α ought to be rather independent of the nature of R_1 but somewhat dependent on the nature of Ar and R_2 (i.e., on carboxonium ion stability) because of ion pairing forces within the (carboxonium ion)·(A⁻)·(R₁OH) aggregate. Thus increased carboxonium ion stability produces a large rate increase accompanied by a slight decrease in the Brønsted α when diffusional separation is rate determining, whereas decreased nucleophilicity in R_1OH results in a change in rate-determining step from k_2 to k_1 , which produces a significant decrease in the Brønsted α .

It bears noting that, if the series $ArCH(OEt)_2$ hydrolyzes increasingly with rate-limiting diffusional separation of the aggregate (v_2 in eq 4) as Ar becomes electron withdrawing, then almost certainly the simple alkyl acetals of aliphatic aldehydes and ketones do also; that is, eq 4 may be the "normal" or "typical" mechanism of acetal hydrolysis.

An interesting development arises from a different presentation of the data in Table II, as shown in Figure 5 as a plot of k_{HA} vs. σ^* for the series of buffer acids. Note that the curve passes through a minimum and that the position of the minimum shifts toward a larger σ^* as pK_{HA} decreases (cf. Table II data). This could be the result of $\partial\alpha/\partial\sigma^*$ or $\partial\rho^*/\partial pK_{HA}$

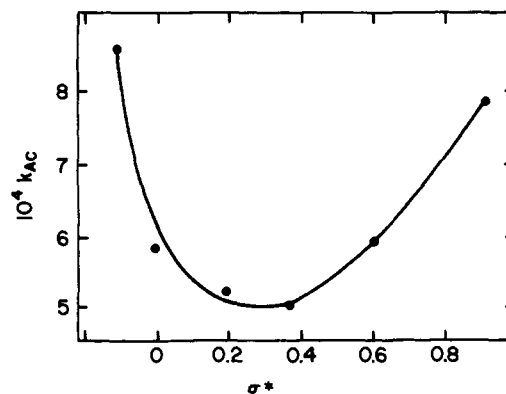


Figure 5. A plot of k_{ac} vs. σ^* for the acetic acid catalyzed hydrolysis of benzaldehyde *O*-ethyl *O*-alkyl acetals.

$\neq 0$ (alluded to earlier), where in this case ρ^* for k_{H^+} is sufficiently small ($\rho^* = -1.07$, correlation coefficient = 0.991) so as to allow the inversion of the substituent effect to be observable for moderately weak acids. In a practical sense, this means that k_{HA} increases as R_1OH becomes a better leaving group if the catalyzing acid is sufficiently weak. This could be a result of ③ and ④ motion in Figure 3, which eventually makes the R_1O moiety partially negative in the transition state when R_1 contains electron withdrawing groups and the catalyzing acid is moderately weak. (It is no coincidence that this describes the natural substrates and active site catalysts for the glucosyl transferases.¹⁸)

Alternatively (and preferably), the minimum in Figure 5 can be viewed as a result of a change in rate-determining step in eq 4: the rate decrease with increasing σ^* on the left "leg" of the curve, where v_2 is rate determining, reflects a commensurate decrease in k_1 and the rate increase with increasing σ^* on the right "leg" of the curve, where v_1 is rate determining, reflects a commensurate increase in k_1 . That the left leg is of steeper slope than the right leg is a result of the increase in k_1 being much less dependent on σ^* than the inverse dependence of k_1 . This is true because of the location of the transition state in Figure 3 for weak catalyzing acids, ③ and ④, and the steeper right "leg" for k_{cac} compared to k_{ac} confirms this motion. (Aside from the points mentioned earlier, another good reason to prefer the rate-limiting diffusional separation alternative is that for the minimum to arise from a $\partial\rho^*/\partial pK_{HA}$ effect requires ρ^* to decrease with increasing pK_{HA} : we observe essentially no effect on ρ^* of pK_{HA} for rate-determining v_2 processes. This is best represented by the several data points in Figure 4 showing $\partial\alpha/\partial\sigma^* \approx 0$, which requires that $\partial\rho^*/\partial pK_{HA} \approx 0$.)

Finally, a brief analysis of the $ArCH(OR)_2$ series, which combines the variables (a) and (c) above, is presented. In this case $R_1 = R_2$ and as R_2 becomes electron withdrawing (relative to ethyl) it destabilizes the carboxonium ion just as does an electron-withdrawing aryl group. The aryl group apparently compensates for this increased electronic demand by increased conjugative stabilization: for $ArCH(OEt)_2$ $\rho = -3.16$ and $\rho^r = -0.29$ and for $ArCH(OCH_2CH_2Cl)_2$ $\rho = -3.20$ and $\rho^r = -1.3$. More generally, the data in Table I show that when $R_1 = R_2$ contains electronegative groups the reactivity diminishes and α decreases (which again shows that when the leaving group (R_1) is better, the predominant effect is reflected in α , consistent with the mechanism of eq 4). For a specific case, the di-2-chloroethyl acetal of benzaldehyde reacts 0.08 times as fast as the mixed acetal, benzaldehyde 2-chloroethyl ethyl acetal (k_{H^+} values), but the α values are indistinguishable.

Solvent Effects. As tabulated in Table III, the rate of hydrolysis falls off markedly as the percent dioxane in the

aqueous solvent is increased ($\mu = 0.5$ with KCl) and ρ increases from 3.16 in H₂O to 3.58 in 50% dioxane. Also, the resonance stabilization factor ρ^r (as measured by the two-parameter Yukawa-Tsuno equation)⁶ increases from -0.29 in water to -0.93 in 50% dioxane. This is precisely as expected from the mechanism of eq 4; as the dielectric constant of the solvent is lowered and the activity of water decreased (and its ability to stabilize ions by solvation), v_{-1} increases relative to v_2 and rate-determining diffusional separation will become more important. Since in water we propose v_2 rate limiting for those acetals less reactive than benzaldehyde diethyl acetal, in 50% dioxane ρ likewise ought to reflect a transition state with a fully formed carboxonium ion. (Because of the nature of the two-parameter $\sigma\rho$ treatment, all the ρ values represent the substituent effect on the acetal going to a fully formed carboxonium ion transition state). The increasing ρ and ρ^r as percent dioxane is increased is a result of increasing electronic demand on the substituents, brought about by decreased solvation of the carboxonium ion by the solvent.

Enzyme Catalysis. These results provide evidence illuminating two features of the chemistry occurring at the active site of lysozyme.¹⁸ First, Asp-52 is present at the active site in the ionized state: our results provide an important role for Asp-52 which need not show up as a high rate acceleration in model systems. Namely, Asp-52 can behave as A⁻ in the mechanism of eq 4. This may well slow down the collapse of the aggregate by a v_{-1} process and thus favor the leaving group diffusing away. In the absence of any evidence for a covalently bound intermediate (similar to an acylal), this carboxonium ion stabilization is best viewed as electrostatic stabilization (much as originally proposed by Phillips).^{18c} Thus, this work provides experimental corroboration of an earlier hypothesis.

The second insight arises from the discussion of Figure 5 and, simply stated, says that the rate of catalysis by Glu-35 of the hydrolysis of the natural substrate is much greater than predicted from previous physical organic models, since the leaving group substituent effect for such a weak catalyzing acid is actually opposite to that of proton catalysis. In this regard, we now are in the process of quantitating this reversal of substituent effects, having noted that Kankaanpera observed substituent effects similar to Figure 3 for proton-catalyzed hydrolysis of the 2-alkoxydihydropyrans and dihydrofurans.¹⁹ With reference to eq 4, if v_2 is the rate-determining step, then $k_2/k_{-1} < 1.0$ and, provided that Asp-52 can change the rate-determining step to v_1 , the rate will increase substantially; just as significantly, Glu-35 can now function as a kinetically more efficient general acid catalyst (cf. motions ③ and ④, Figure 3). Thus our results would predict that the major role of Asp-52 is to change the rate-determining step from diffusional separation (v_2 in eq 4) to proton transfer concerted with C-O bond cleavage (v_1 , in eq 4). This is a vital role since the rate enhancement caused by omitting the k_2/k_{-1} term is augmented by the greater catalysis by Glu-35 on v_1 . Previous physical organic models have not been able to elucidate these roles because this is the first study reporting such an interplay between pK_{HA} and substituent effects on the leaving group in

systems showing a probable change in rate-determining step.

Semiquantitatively, Figure 5 allows an estimation of the rate of a rate-determining v_1 process relative to the rate of a rate-determining v_2 process for the acetic acid catalyzed hydrolysis of benzaldehyde trifluoroethyl ethyl acetal: assuming $\rho^* = -1.7$ (the slope "established" by the OEt and OCH₃ points in Figure 5) for a rate-determining v_2 process and extrapolating to the trifluoroethyl group ($\sigma^* = 0.92$) produces a factor of 50. That is, if this model is correct, acetic acid functions as a catalyst 50 times more effectively when v_1 is rate determining than when v_2 is rate determining. This factor will increase as the catalyzing acid pK_a increases and as the substituent effect changeover occurs "earlier". Fife has recently reported a rate enhancement of 100-fold for a rate-determining v_1 process.²⁰

Work is in progress on a model which better represents lysozyme substrates and hopefully a more quantitative measurement of this type of rate enhancement will be provided by that system.

Acknowledgment. Support from the National Science Foundation (CHE77-00482) is gratefully acknowledged. Kathleen Cannon synthesized the phosphonate buffers and Raymond Johnson made some of the preliminary kinetic measurements.

References and Notes

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